RNA Purification Magnetic Beads

Cat. No. G971

Store at 4°C.

Name	Quantity	Cat. No.
RNA Purification Magnetic Beads	5ml	G971

Product Description

abm's RNA Purification Magnetic Beads are paramagnetic particles coated with carboxyl groups that can reversibly bind to nucleic acids. The RNA Purification Magnetic Beads are specifically formulated for usage with RNA and can be used to concentrate and purify samples resulting in high purity, reproducibility and yield.

Key Features

- RNase-free.
- Simple automation-friendly protocol with no centrifuge required.
- Complete removal of salts, enzymes and other impurities.
- Cost effective alternative to other commercially available products e.g. RNAClean XP beads.

Protocol

Additional Materials Required (not included)

- Freshly prepared 80% ethanol
- Nuclease-free water
- Magnetic separation rack

Notes: Before starting, prepare 80% ethanol fresh prior to each experiment. This is a standard protocol that is compatible with 1.5ml or 0.2ml tubes.

1. Remove RNA Purification Magnetic Beads from 4°C and incubate at room temperature for 30min.

2. Vortex the RNA Purification Magnetic Beads thoroughly for 30s.

3. Add RNA Purification Magnetic Beads to sample at a 1.8X ratio (e.g. for a 50µl sample, add 90µl of beads) and pipette ten times to thoroughly mix. Incubate at room temperature for 10min.
   •If sample is <50µl, increase the volume to 50µl by adding an appropriate amount of nuclease-free water.

4. Place tube onto magnetic rack for 5min or until the solution becomes clear.

5. Keep the tube on the magnetic rack; carefully remove the supernatant by pipette and discard.

6. Keep the tube on the magnetic rack; add 200µl of freshly prepared 80% ethanol. Dispense the liquid onto the opposite side of the tube in order to avoid disturbing the beads. Incubate 30s and then remove the supernatant by pipette. Repeat for a total of two washes.

7. Keep the tube on the magnetic rack; remove residual ethanol by pipette and then open tube caps to air dry for 2-5min. Monitor the appearance of the beads closely.
   •Beads are ready for elution when they appear "matte." Beads that appear "shiny" still contain residual ethanol; beads that appear "cracked" have been over-dried. The latter two scenarios will result in contamination and reduced yield.

8. Add ≥20µl of nuclease-free water to the RNA Purification Magnetic Beads and remove the tube from the magnetic rack. Pipette the mixture thoroughly to resuspend. Incubate at room temperature for 5min.

9. Place the tube on the magnetic rack for 5min or until the solution becomes clear. Carefully remove the supernatant by pipette and transfer into a new sterile tube.
   •Ensure no bead carry-over. If beads become dislodged, simply mix the sample again and place tube back onto the magnetic rack for separation.

10. The purified RNA is ready for downstream applications or long term storage at -80°C.

General Notes

- Store tightly sealed at 4°C upon arrival. Do not freeze!
- Beads appear brown and may settle during storage. Once thoroughly vortexed it should appear homogenous and consistent in colour.
- Ensure beads come to room temperature (20-25°C) before use; using or storing beads at the incorrect temperature will result in lower yield and difficulty with handling.
- Step 7 and 8 are critical for sample recovery. Some samples may be ready for elution before others; it may be prudent to add nuclease-free water immediately to these samples rather than waiting the full 5min to air dry.

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